

14 September 2020

To: Recipients of M100, 30th ed.  
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 Vice President, Standards and Quality  
 Subject: Combined Corrections

This notice is intended to inform users of corrections made to CLSI document M100, *Performance Standards for Antimicrobial Susceptibility Testing*, 30th ed. The corrections are described below and shown as highlighted and/or stricken text in the table excerpts.

**Correction: 10 September 2020**

**Appendix E. Dosage Regimens Used to Establish Susceptible or Susceptible-Dose Dependent Breakpoints:**

The susceptible minimal inhibitory concentration (MIC) breakpoint for *Pseudomonas aeruginosa* and ceftazidime-avibactam is listed incorrectly as “≥ 8/4 µg/mL.” The susceptible MIC breakpoint has been corrected to read “≤ 8/4 µg/mL” for accuracy and is consistent with Table 2B-1.

**Appendix E. Dosage Regimens Used to Establish Susceptible or Susceptible-Dose Dependent Breakpoints**

Antimicrobial Agent	Breakpoints and Interpretive Categories	
	Susceptible	
	MIC	Dose
<b>Table 2B-1. <i>Pseudomonas aeruginosa</i></b>		
Ceftazidime-avibactam	≥ 8/4 µg/mL ≤ 8/4 µg/mL	2.5 g (2 g ceftazidime + 0.5 g avibactam) administered every 8 h over 2 h

**Correction: 28 April 2020**

**Appendix I. Cefiderocol Broth Preparation and Reading Broth Microdilution Minimal Inhibitory Concentration End Points:**

In section I1.2, in the step-action table describing preparation of zinc stock solution, the comment for step 1 is listed incorrectly as, “This solution contains 10 mg Zn<sup>++</sup>/mL.” The comment has been corrected to read, “This solution contains 0.65 mg Zn<sup>++</sup>/mL (10 mmol Zn<sup>++</sup>/mL).”

Step	Action	Comment
1	Dissolve 0.29 g ZnSO <sub>4</sub> • 7H <sub>2</sub> O in 100 mL deionized water.	This solution contains 400.65 mg Zn <sup>++</sup> /mL (10 mmol Zn <sup>++</sup> /mL).
2	Sterilize the solution by membrane filtration.	
3	Store the solution at 15 to 25°C.	

In section I2, in the step-action table showing an example for preparing cation-adjusted Mueller-Hinton broth that contains below-detectable concentrations (< 0.0001 mg/L) of Zn<sup>++</sup> after chelation:

- Step 2 incorrectly reads, “Add 0.1 mL Zn<sup>++</sup> stock per L to obtain a concentration of 1 mg/L.” Step 2 has been corrected to read, “Add 1.54 mL Zn<sup>++</sup> stock per L (1.54 mL for each 1 mg/L).”
- The comment for step 2 is listed incorrectly as “1 mg/mL • 0.1 mL = 0.1 mL.” The comment has been corrected to read, “1 mg/L • 1.54 mL (0.65 mg Zn<sup>++</sup>/mL) = 1 mg/L”.

Step	Action	Comment
1	Calculate the amount of Zn <sup>++</sup> needed using this formula:  Final amount needed – amount in medium = amount to be added	For Zn <sup>++</sup> , the final amount needed is 0.5-1 mg/L.  1 mg/L – 0 mg/L = 1 mg/L
2	Add <del>0.1</del> 1.54 mL Zn <sup>++</sup> stock per L (1.54 mL for each 1 mg/L). <del>to obtain a concentration of 1 mg/L</del>	1 mg/mL • <del>0.1</del> 1.54 mL (0.65 mg Zn <sup>++</sup> /mL) = <del>0.1</del> 1 mg/L
3	Proceed with steps 8 and 9 above.	

Correction: 6 February 2020

Table 2C. Zone Diameter and MIC Breakpoints for *Staphylococcus* spp.:

In general comment (5), in the Methods for Detection of Methicillin (Oxacillin)-Resistant *Staphylococcus* spp. table, the incubation period for *Staphylococcus epidermidis* with cefoxitin disk diffusion is listed incorrectly as “16-18 h.” The incubation period has been corrected to read “24 h.”

- (5) Most methicillin (oxacillin) resistance is mediated by *mecA*, encoding PBP2a (also called PBP2'). Isolates that test positive for *mecA* or PBP2a should be reported as methicillin (oxacillin) resistant (see Appendix H).

Detection of methicillin (oxacillin) resistance in staphylococci is achieved by using specific methods as listed in Table 2C and further described in Table 3F.

Organism	Methods for Detection of Methicillin (Oxacillin)-Resistant <i>Staphylococcus</i> spp.				
	Cefoxitin MIC	Cefoxitin disk diffusion	Oxacillin MIC	Oxacillin disk diffusion	Oxacillin salt agar
<i>S. aureus</i>	Yes (16-20 h)	Yes (16-18 h)	Yes (24 h)	No	Yes (24 h)
<i>S. lugdunensis</i>	Yes (16-20 h)	Yes (16-18 h)	Yes (24 h)	No	No
<i>S. epidermidis</i>	No	Yes (16-18 h)	Yes (24 h)	Yes (16-18 h)	No
<i>S. pseudintermedius</i>	No	No	Yes (24 h)	Yes (16-18 h)	No
<i>S. schleiferi</i>	No	No	Yes (24 h)	Yes (16-18 h)	No
Other <i>Staphylococcus</i> spp. (not listed above)	No	Yes <sup>a</sup> (24 h)	Yes <sup>a</sup> (24 h)	No	No

Abbreviations: h, hour(s); MIC, minimal inhibitory concentration; MRS, methicillin (oxacillin)-resistant staphylococci; PBP2a, penicillin-binding protein 2a.

<sup>a</sup> For isolates of “other *Staphylococcus* spp.” from serious infections for which the oxacillin MICs are 0.5-2 µg/mL, testing for *mecA* or PBP2a should be considered (see comment [17]). Cefoxitin disk diffusion is not currently recommended.

Mechanisms of methicillin (oxacillin) resistance other than *mecA* are rare and include a novel *mecA* homologue, *mecC*. MICs for strains with *mecC* are typically cefoxitin resistant and oxacillin susceptible; *mecC* resistance cannot be detected by tests directed at *mecA* or PBP2a.

If you require any additional clarification regarding these corrections, please contact CLSI Customer Service ([customerservice@clsi.org](mailto:customerservice@clsi.org)).

We appreciate your commitment to CLSI and regret any inconvenience.